Large Animal Specialist products

Laparoscopic Set for repositioning and fixation of abomasal displacement in Cattle.

Dr. Fritz offers a new set for the correction of abomasal displacement in dairy cattle using a minimally invasive technique developed by Dr. Janowitz. This set allows the veterinary surgeon to use a quick and extremely safe technique with no medication requirements as well as quick recovery of the patient. The procedure can be carried out in the field. The equipment is easy to handle and to clean. At the heart of the set is a specially developed robust scope with a length designed specifically for this purpose.

Instruments are specifically designed for use in large animal field situations and stored in a practical transport case to guarantee a long life.

In research programs it emerged that the traditional toggle system often offered inadequate strength and resulted in either rupture of the suture material or breakage of the plastic toggle material. This has led to the development of a new safety toggle with a 90 cm long break safe double suture.

Set consists of:
- Wide Angled Telescope
- LED Battery Light Handle and Spare Bulb
- Suture Grasping Forceps, long
- Inflation and Puncture Needle
- Trochars and Trochar Cannulas
- Abomasal Puncture System
- Safety Toggle Pins (5 pcs)
- Christiansen’s Spieker Insertion Tool and Trochar
- Sterilising and Storage Tray
- Inflation Pump
- Cleaning Brushes
- Transportation Case

A range of accessories have been also developed by Dr Christiansen to enable laproscopic repositioning in a standing upright position.

For further information regarding the technique and detailed information about the set, please visit www.vetinst.com or request the Dr Fritz brochure on Bovine Laparoscopy. (Code: BRCOW)

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RecSpec™
RAMALT (Recto-anal mucosa-associated lymphoid tissue) Speculum. www.ramalt.com

Eradication of TSE infections is a long standing goal for the UK policy on animal health and consumer protection. The national scrapie plan (NSP) aims to reduce the incidence of scrapie within the National Flock by breeding for genetic resistance to the disease. In addition, control of scrapie is facilitated by the elimination of clinical scrapie suspect cases and by management procedures directed to block or minimise transmission of the infection. Moreover, detection and subsequent removal of pre-clinical scrapie cases would help preventing the spread of the infection, both within and between flocks, and would facilitate epidemiological monitoring, if the technique(s) involved were suitable for high throughput sampling.

The tissue source should be both rich in disease-specific prion protein (PrPd) and accessible as a field biopsy. Two tissues amenable to biopsy are known to contain diagnostically useful amounts of the disease-specific prion protein (PrPd). However, both palatine tonsil and third eyelid biopsies are difficult to obtain, requiring total anaesthesia and experience to be performed accurately, and none of them are likely to have practical field application. A further source, the rectal mucosa at the recto-anal junction contains abundant lymphoid tissue (RAMALT) in an anatomically accessible site, and in preliminary studies carried out on clinically affected sheep, it has been shown that it is also strongly positive for PrPd. It is also easily sampled using simple biopsy instruments and does not require anaesthesia, being in principle suitable for large scale field studies. The technique is greatly simplified by use of a purpose designed speculum and a disposable scissor and forceps

Materials & Methods
The RAMALT is located immediately beneath the last 1cm of rectal mucosa at the end of the alimentary tract. The lymphoid follicles are found in aggregates underneath the crypts of the rectal mucosa folds (Fig 1) and become more visible when treated with glacial acetic acid (Fig. 2) and trans-illuminated (Fig. 3). This information was vital for obtaining the correct material in biopsies from live animals (Fig. 4).

Biopsy Procedure in Live Animals
We have developed a biopsy procedure to obtain RAMALT tissue from live sheep including the design, manufacture and testing of a disposable speculum to gain access to the target 1cm² area of rectal mucosa (Fig. 5). The speculum is inserted in the anus after liberal application of analgesic cream which also doubles as a lubricant. The folds of the rectal mucosa are now easily identified (Figs. 6 & 7).

- A sample of RAMALT tissue is taken by picking up the base of a crypt with forceps and cutting off the resultant ‘tent’ of tissue with scissors (Fig. 8).

- This precise location of sampling ensures adequate RAMALT lymphoid follicles are present that is essential for the detection of scrapie.¹

- Biopsy sample fixed and processed, thin sections cut and mounted on glass slides. Tissue sections examined for the presence of the disease associated marker PrPd (Fig. 9).²

- Brown staining within the lymphoid follicles is a positive result for scrapie (Fig. 9). The same would apply for deer with CWD.

The ramalt speculum can be purchased in packs of 10 or 100. A single use disposable set of scissors and forceps are also available in 10’s or 100’s.

References and acknowledgements

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